

Polydatin attenuated food allergy *via* store-operated calcium channels in mast cell

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Abstract

AIM: To investigate the effect of polydatin (PD), a resveratrol glucoside, on mast cell degranulation and anti-allergic activity.

METHODS: After the rats were orally sensitized with ovalbumin (OVA) for 48 d and underwent PD treatment for 4 d, all the rats were stimulated by 100 mg/mL OVA for

24 h and then sacrificed for the following experiments. The small intestines from all the groups were prepared for morphology examination by hematoxylin and eosin staining. We also used a smooth muscle organ bath to evaluate the motility of the small intestines. The OVA-specific immunoglobulin E (IgE) production and interleukin-4 (IL-4) levels in serum or supernatant of intestinal mucosa homogenates were analyzed by enzyme-linked immunosorbent assay (ELISA). Using toluidine blue stain, the activation and degranulation of isolated rat peritoneal mast cells (RPMCs) were analyzed. Release of histamine from RPMCs was measured by ELISA, and regulation of PD on intracellular Ca^{2+} mobilization was investigated by probing intracellular Ca^{2+} with fluo-4 fluorescent dye, with the signal recorded and analyzed.

RESULTS: We found that intragastric treatment with PD significantly reduced loss of mucosal barrier integrity in the small intestine. However, OVA-sensitization caused significant hyperactivity in the small intestine of allergic rats, which was attenuated by PD administration by 42% (1.26 ± 0.13 g *vs* OVA 2.18 ± 0.21 g, $P < 0.01$). PD therapy also inhibited IgE production (3.95 ± 0.53 ng/mL *vs* OVA 4.53 ± 0.52 ng/mL, $P < 0.05$) by suppressing the secretion of Th2-type cytokine, IL-4, by 34% (38.58 ± 4.41 pg/mL *vs* OVA 58.15 ± 6.24 pg/mL, $P < 0.01$). The ratio of degranulated mast cells, as indicated by vehicles (at least five) around the cells, dramatically increased in the OVA group by 5.5 fold ($63.50\% \pm 15.51\%$ *vs* phosphate-buffered saline $11.15\% \pm 8.26\%$, $P < 0.001$) and fell by 65% after PD treatment ($21.95\% \pm 4.37\%$ *vs* OVA $63.50\% \pm 15.51\%$, $P < 0.001$). PD mediated attenuation of mast cell degranulation was further confirmed by decreased histamine levels in both serum (5.98 ± 0.17 *vs* OVA 6.67 ± 0.12 , $P < 0.05$) and intestinal mucosa homogenates (5.83 ± 0.91 *vs* OVA 7.35 ± 0.97 , $P < 0.05$). Furthermore, we demonstrated that administration with PD significantly decreased mast cell degranulation due to reduced Ca^{2+} influx through store-operated calcium channels (SOCs) (2.35 ± 0.39 *vs* OVA 3.51 ± 0.38 , $P < 0.01$).

CONCLUSION: Taken together, our data indicate that PD stabilizes mast cells by suppressing intracellular Ca^{2+} mobilization, mainly through inhibiting Ca^{2+} entry *via* SOCs, thus exerting a protective role against OVA-sensitized food allergy.

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Key words: Polydatin; Food allergy; Mast cells; Store-operated calcium channels; Ca^{2+}

Core tip: In the present study, we have demonstrated for the first time that polydatin has the capacity for preventing pathogenesis of food allergy, which is dependent on regulation of Ca^{2+} mobilization *via* store-operated calcium channels in mast cells.

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INTRODUCTION

Food allergy (FA) is an adverse reaction mediated by immunoglobulin E (IgE) or non-IgE antibodies^[1], which involves an abnormal response by the immune system to specific proteins in foods^[2]. FA has been recognized as a worldwide health problem, especially in western countries, which is due to the severity of the reactions and its dramatic increase over the past three decades^[3-5]. The majority of food allergies worldwide are caused by “eight main food allergens”, including peanuts, tree nuts, eggs, milk, fish, crustacean shellfish, wheat, and soy^[6]. It has been suggested that 25% of infants^[7], 8% of children^[3-5], and 2%-5% of adults^[8] suffer from FA. However, the current understanding about the etiology of food allergies remains poor, and no effective treatment is available except the preventative measure of avoiding the offending food in the diet.

Mast cells play an essential role in the development of intestinal inflammatory disorders during food allergy. Cross-linking of the high-affinity IgE receptor (FcεRI) on mast cells by allergens results in degranulation, leukotriene generation, and cytokine synthesis. Degranulated mast cells release inflammatory mediators, including histamine and Th2 cytokines^[9], which cause abnormal gut contractions and intestinal mucosa damage, which in turn result in abdominal pain, cramps, vomiting, and/or diarrhea^[10]. It has been known that IgE-dependent mast cell degranulation relies on intracellular Ca^{2+} signaling^[11,12]. Cytoplasmic Ca^{2+} mainly comes from the stored Ca^{2+} in the endoplasmic reticulum (ER) and extracellular Ca^{2+} through store-operated calcium channels (SOCs)^[13]. Therefore, modulation of Ca^{2+} mobilization is

a potential therapeutic strategy for stabilizing mast cells upon FcεRI activation and potentially offering a novel treatment method for allergic diseases.

Polydatin (PD), also known as polygona cuspidati radix, is a natural component isolated from *Polygonum cuspidatum*. It has been determined as a resveratrol glucoside with a 3,4,5-trihydroxystilben-3-*D*-mono-*D*-glucoside molecular structure. Previous studies have demonstrated that PD has a therapeutic effect on the treatment of allergic diseases. Using the passive cutaneous anaphylaxis (PCA) model in mice, Lim *et al.*^[14] showed that PD reduced mast cell degranulation by suppressing phosphorylation of Syk and mitogen-activated protein kinases. On the other hand, Yuan *et al.*^[15] found that PD alleviated PCA in mice by stabilizing mast cells *via* the inhibition of Ca^{2+} release-activated Ca^{2+} channels. However, the therapeutic effect of PD on food allergies has not yet been determined.

In this study, we established ovalbumin (OVA)-induced food allergic models and evaluated the therapeutic effect of PD on food allergy. Furthermore, we explored the effect of PD on mast cell degranulation and found that the underlying mechanism was related to Ca^{2+} mobilization. Our research presented here is the first to reveal that PD can inhibit food allergy by suppressing mast cell degranulation *via* regulation of SOCs.

MATERIALS AND METHODS

Animals

Four-week old female Brown-Norway (BN) rats were purchased from Vital River Laboratories (Beijing, China) and housed in groups of four per cage in a controlled environment with a photoperiod of 12 h light to 12 h dark and a temperature of 20 ± 2 °C. Sanitary controls were performed for all major rodent pathogens, with the results of these tests being uniformly negative. All the animal experimental procedures were approved by the Animal Care and Use Committee of Shenzhen University and carried out in accordance with the Guide for the Care and Use of Laboratory Animals published by the United States National Institutes of Health (publication No. 85-23, revised 1996).

Forty-eight Brown-Norway rats were randomly divided into four groups: control group ($n = 12$), OVA group ($n = 12$), OVA + PD group ($n = 12$), and PD group ($n = 12$). Each group received phosphate-buffered saline (PBS), OVA, or PD, as shown in Figure 1^[16]. The control group received 1 mL PBS (0.1 mol/L) daily by gavage administration for 52 d, while the OVA group was orally treated with 1 mg OVA (1 mg/mL) for the first 48 d and 1 mL PBS (0.1 mol/L) from days 48 to 52. The OVA + PD group received PD (150 mg/mL \times 1 mL daily per rat) oral treatment daily from days 49 to 52 after OVA sensitization. The PD group was not challenged by OVA. All the groups of rats were stimulated by 100 mg/mL OVA for 24 h at the end of day 52 and then sacrificed for the following experiments.

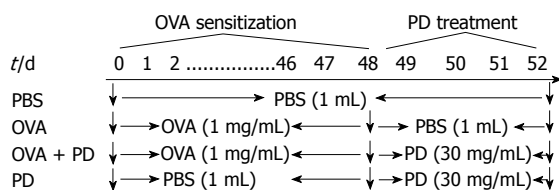


Figure 1 Protocol of ovalbumin sensitization/challenge and Formula-3 treatment. Rats were sensitized with ovalbumin (OVA) (1 mg/mL × 1 mL daily per rat) intragastrically for 48 d. For the polydatin (PD) treatment group, the rats were orally treated with PD (30 mg/mL × 1 mL daily per rat) from days 49 to 52 after OVA sensitization. All the rat groups were challenged by 100 mg/mL OVA for 24 h at the end of day 52 and then sacrificed. PBS: Phosphate-buffered saline.

Hematoxylin and eosin staining in small intestine tissues

The jejunal parts of the small intestine were isolated from the rats and embedded with paraffin. Sections (7 μm) were prepared and subjected to hematoxylin and eosin (HE) staining as previously reported^[17].

Measurements of smooth muscle contractility

The tension of smooth muscle contractility was measured as previously reported^[18]. Briefly, 2 cm long segments of the small intestine from the upper part of the jejunum to the lower part of the ileum were cut and mounted by hanging from triangle hooks. The hooks were connected to transducers from the upper end, and were inserted through the gut lumen from the lower end, allowing the circular muscle to contract. The tissue segments were incubated in chambers containing 20 mL Tyrode's solution (136 mmol/L NaCl, 5.4 mmol/L KCl, 1.0 mmol/L MgCl₂, 0.33 mmol/L NaH₂PO₄, 1.8 mmol/L NaCl, 10.0 mmol/L glucose and 5.0 mmol/L HEPES), which was kept at 37 °C and constantly aerated with a mixture of 95% oxygen and 5% carbon dioxide. The initial tension load was set at 1.0 g, from which the segments spontaneously relaxed over time. The segments were allowed to stabilize for 30 min before they were stimulated with 1 mg/mL OVA for 3 min. *R0* was defined as the mean basal tension, when the segments were under rest conditions. ΔR denotes (*R1-R0*), where *R1* is the contract tension when the segments were stimulated with OVA.

Toluidine blue stain

Typical mast cells in rat small intestine tissue or peritoneal lavage solution (RPLS) were stained with toluidine blue stain as previously described^[19]. Briefly, 200 L RPLS was air dried on cromolyn sodium pretreated slides and then covered with several drops of staining solution (toluidine blue stain dissolved in 70% ethanol). After 90 s, the staining solutions were washed away quickly with running tap water and the stained cells were examined and counted under a light microscope (Olympus, Japan).

Enzyme-linked immunosorbent assay

The contents of interleukin-4 (IL-4) (eBioscience Inc.,

CA, United States) and histamine (R & D Inc., MN, United States) in RPLS and serum were assayed by commercial enzyme-linked immunosorbent assay (ELISA) kits using paired antibodies according to the manufacturer's instructions. Serum IgE levels were also checked using a commercial ELISA kit (BD Pharmingen, CA, United States), following the manufacturer's instructions.

Rat peritoneal mast cell isolation

The BN rats were sacrificed after being anaesthetized by ether inhalation in air. Rat peritoneal mast cells (RPMCs) were obtained by peritoneal lavage and purified by density gradient fractionation as described previously^[20,21]. Isolated RPMCs preparations contained > 98% mast cells and at least 98% of these cells were viable, as checked by metachromatic staining in 0.05% toluidine blue.

Ca²⁺ imaging by confocal microscope

Intracellular Ca²⁺ signal was measured as described previously with minor modification^[22]. RPMCs or RBL-2H3 cells were incubated with 5 μmol/L Ca²⁺ fluorescent probe fluo-4 AM (Invitrogen, CA, United States) for 30 min at room temperature. After washing with Tyrode's solution three times, the dye inside the cells was allowed to de-esterify for 30 min at 37 °C. It has been determined that nearly 95% of the fluorescent dye was retained in the cytoplasm. Fluorescent images of Ca²⁺ were obtained using an Olympus 1000 confocal microscope with a 40 × oil immersion lens (NA 1.3) (Olympus, Japan). The fluo-4 signal was excited at 488-nm and emitted at > 505 nm. Frame-scan images were acquired at a sampling rate of 15 ms per frame and 20 s per interval.

Image data were analyzed off-line using fv10-asw.2.1 software. A selected image from each image set was used as a template for designating the region of interest (ROI) within each cell. The integrated intracellular Ca²⁺ concentration was determined by calculating $\Delta F/F0$. *F0* was defined as the mean basal fluorescence intensity of the dye recorded during the first 5-10 scanning frames, when the cells were under rest conditions. ΔF denotes (*F-F0*), where *F* is the temporal fluorescence intensity. The $\Delta F/F0$ values within each ROI were plotted as a function of time (typical time-courses of Ca²⁺ response to thapsigargin or DNP-BSA stimulation in single RBL-2H3 cells). The amplitude of the Ca²⁺ response within each cell was quantified as the highest $\Delta F/F0$ level reached during the measurement period, which was averaged over all cells within each group.

Statistical analysis

Data are presented as mean ± SE. When two comparisons were obtained, Student's unpaired two tailed *t* test was used. When multiple comparisons were obtained, the analyses consisted of one-way analysis of variance for repeated measures and Student-Newman-Keuls multiple comparison test. A value of *P* < 0.05 was considered to be statistically significant.

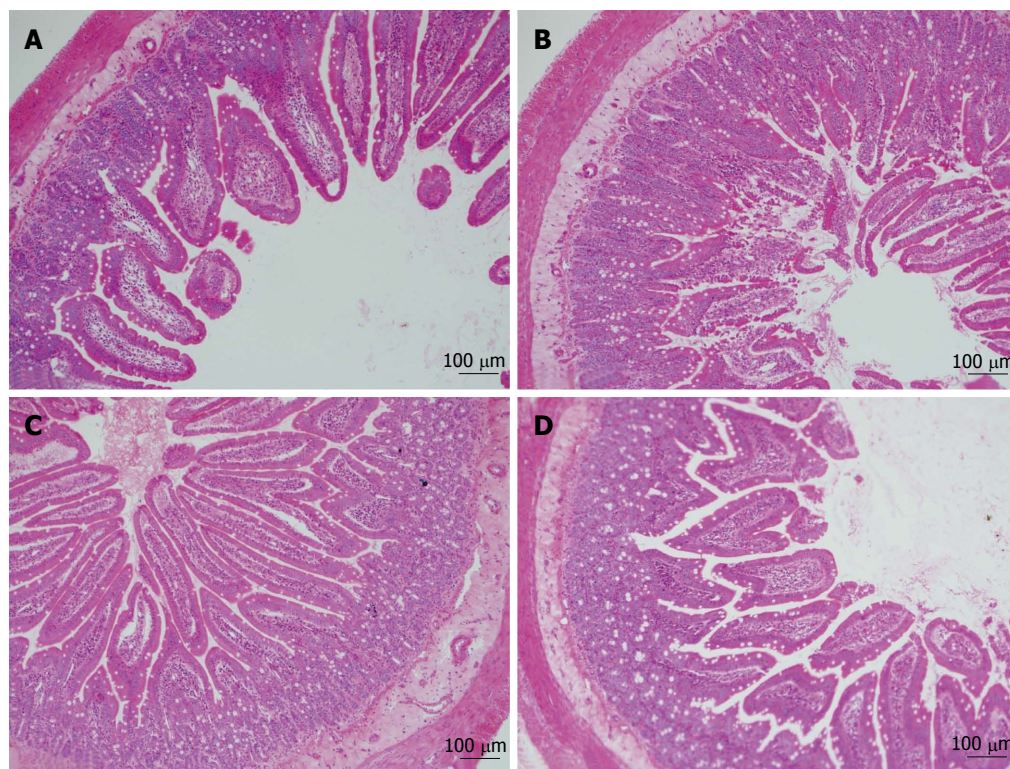


Figure 2 Polydatin attenuated tissue injury in small intestine caused by ovalbumin sensitization. A: Phosphate-buffered saline group; B: Ovalbumin (OVA) group; C: OVA + polydatin (PD) group; D: PD group. Morphology of intestinal jejunum was analyzed by hematoxylin and eosin staining. Representative images from three independent experiments are shown (magnification, $\times 63$).

RESULTS

PD attenuated OVA-challenge caused small intestine abnormality in rats

In the present study, 1 mg OVA was used to sensitize BN rats orally and establish a food allergy model as previously described^[16,23]. Loss of mucosal barrier integrity is a leading cause of food allergy^[24]. Thus, we isolated jejunal fractions from the small intestine and checked tissue damage by HE staining. As shown in Figure 2, the results revealed that the intestinal mucosae were severely injured in the OVA group: the intestinal villi were eroded, and the swelling, shedding, and numbers of intestinal villi were significantly reduced. The morphological abnormality of the small intestine caused by OVA-sensitization was significantly attenuated by PD treatment.

The correlation between intestinal allergy and smooth muscle motility has been indicated by the fact that exposure to luminal allergen induces a state of proximal small intestinal hyperreactivity^[25,26]. In order to evaluate the effects of PD on intestine motility, 2 cm intestine segments from each group were prepared, and the intestinal contraction tension was detected by smooth muscle organ bath. In response to 1 mg/mL OVA, intestinal segments isolated from OVA-allergic rats had a significant higher tension than the PBS group. The elevation of tissue tension caused by OVA sensitization was significantly attenuated by PD treatment by approximately 42% (Figure 3).

Treatment with PD decreased IL-4 levels and attenuated IgE production in OVA-sensitized group

The body weight of all rats in each group was monitored on days 0, 48 and 52. We found that basal body weight

levels on day 0 were similar in all four groups. Compared to the PBS group, OVA sensitization significantly reduced body weight on day 48 (Figure 4A, left panel). After being treated with PD for 4 d, there was no significant difference between the PBS and OVA + PD groups (Figure 4A, left panel), which indicates that PD administration could maintain the body weight of an allergic rat at a normal level. The cytokine levels in the supernatant of the intestine mucosa were measured by ELISA. The results showed that the concentration of IL-4 in the OVA-challenged group was significantly higher than in the control group (58.15 ± 6.24 pg/mL *vs* 35.51 ± 5.48 pg/mL) (Figure 4B). Treatment with PD reduced the enhancement of IL-4 by 34%. Meanwhile, ELISA analysis showed that the concentration of OVA-specific IgE in serum was enhanced by 1.2 fold in the OVA group and PD therapy returned it to a normal level (Figure 4C).

PD reduced mast cell activation and degranulation in small intestine

Mast cell degranulation and histamine release are major factors in food allergy. The number and morphology of the mast cells in rat small intestine tissues (data not shown) or RPLS were examined by toluidine blue stain. In the OVA group, the number of mast cells was significantly increased and the cell size was much bigger, with more shrink on the cell membrane, bubbles in the cytoplasm, and degranulation vehicles around the cells (Figure 5A-D). *In vivo* administration with PD for 4 d reversed OVA-challenge-induced damage in mast cells. The ratio of degranulated mast cell, as indicated by vehicles (at least five) around the cells, dramatically increased in the OVA group by 5.5 fold and fell by 65% after PD treatment

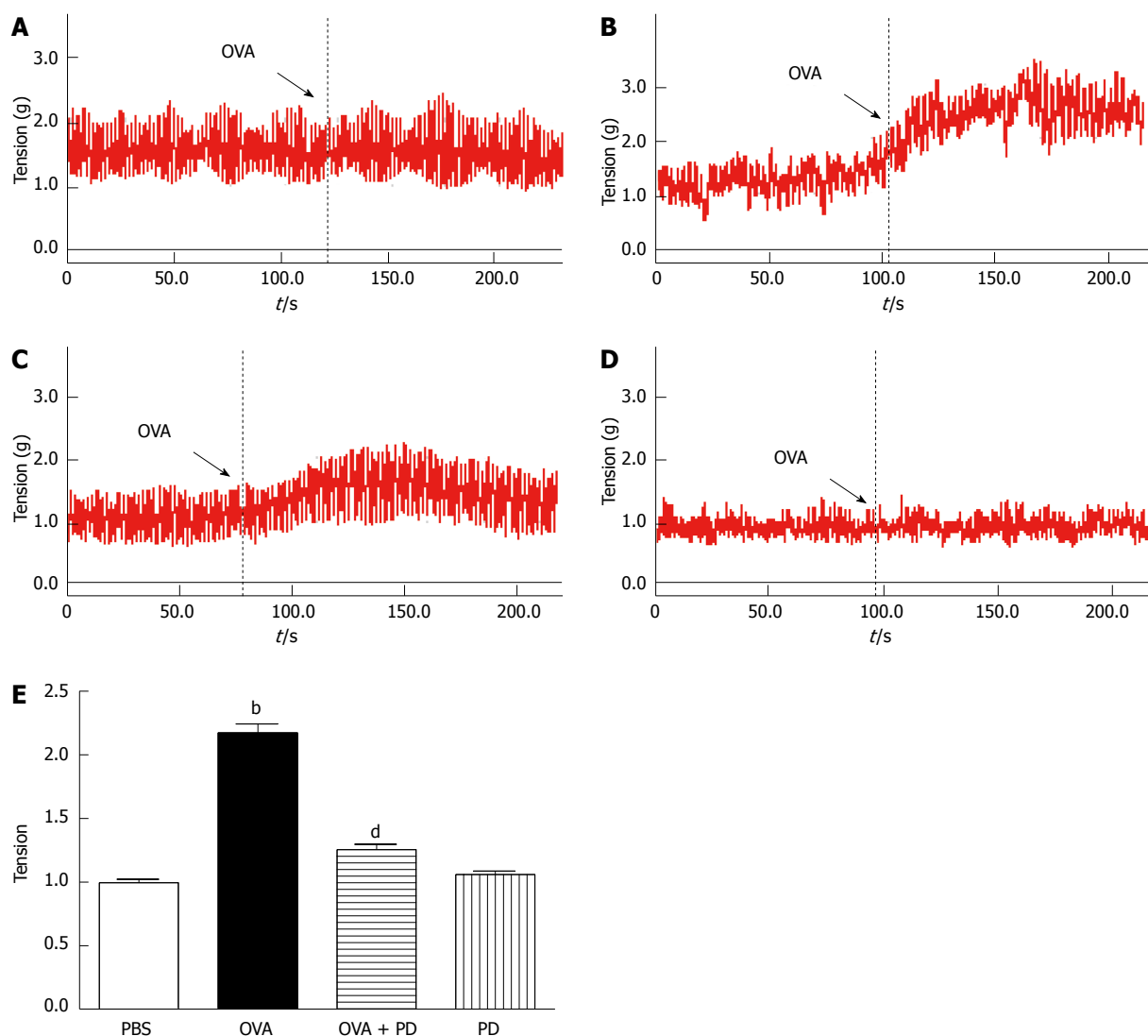


Figure 3 Polydatin attenuated small intestinal hyperreactivity in ovalbumin-allergic rats. A: Phosphate-buffered saline (PBS) group; B: Ovalbumin (OVA) group; C: OVA + polydatin (PD) group; D: PD group. *n* = 8, ^b*P* < 0.01 vs PBS group; ^d*P* < 0.01 vs OVA group. The tension of intestinal mobility was measured by smooth muscle organ bath, typical contraction curves (A-D), and the peak tension of each group (E).

(Figure 5E). PD mediated attenuation of mast cell degranulation was further confirmed by decreased histamine levels. It was found that histamine release in both serum and RPLS was significantly increased in the OVA-induced food allergic group, which was attenuated by PD therapy by approximately 11% and 20% respectively (Figure 5F).

PD inhibited mast cell degranulation by modulating Ca²⁺ mobilization through SOC channels

Rapid translocation of Ca²⁺ has been well-known to be essential for mast cell degranulation^[27]. In a food allergic model, we also found that mast cell activation is related to stimulation of Ca²⁺ mobilization (data not published). To explore the underlying mechanism for the inhibitory effect of PD on mast cell degranulation, we isolated RPMCs and monitored intracellular Ca²⁺ with fluo-4 (5 mol/L). Using a standard Ca²⁺ add-back assay, in which intracellular Ca²⁺ stores were depleted by thapsigargin (TG), a sarcoplasmic/endoplasmic reticulum calcium

ATPase (Ca²⁺ pump) blocker, in Ca²⁺-free extracellular solution, after which the extracellular Ca²⁺ concentration was returned to 2 mmol/L^[28]. Using this protocol, TG elicited two Ca²⁺ peaks, where the first one represented the ER Ca²⁺ release, and the second represented Ca²⁺ entry through activated SOCs. As shown in the first peaks in Figure 6A-D, when the cells were in Ca²⁺-free solution, the TG-evoked Ca²⁺ amplitude was similar in all the groups, suggesting the amounts of Ca²⁺ released from ER are nearly the same. In the presence of 2 mmol/L extracellular Ca²⁺, the TG-evoked Ca²⁺ influx was dramatically enhanced in OVA-sensitized RPMC by 1.5 fold, while PD treatment reduced the Ca²⁺ entry to normal level. The results indicate that PD attenuated OVA-induced Ca²⁺ influx elevation through SOCs.

DISCUSSION

The effect of resveratrol, a structural and functional

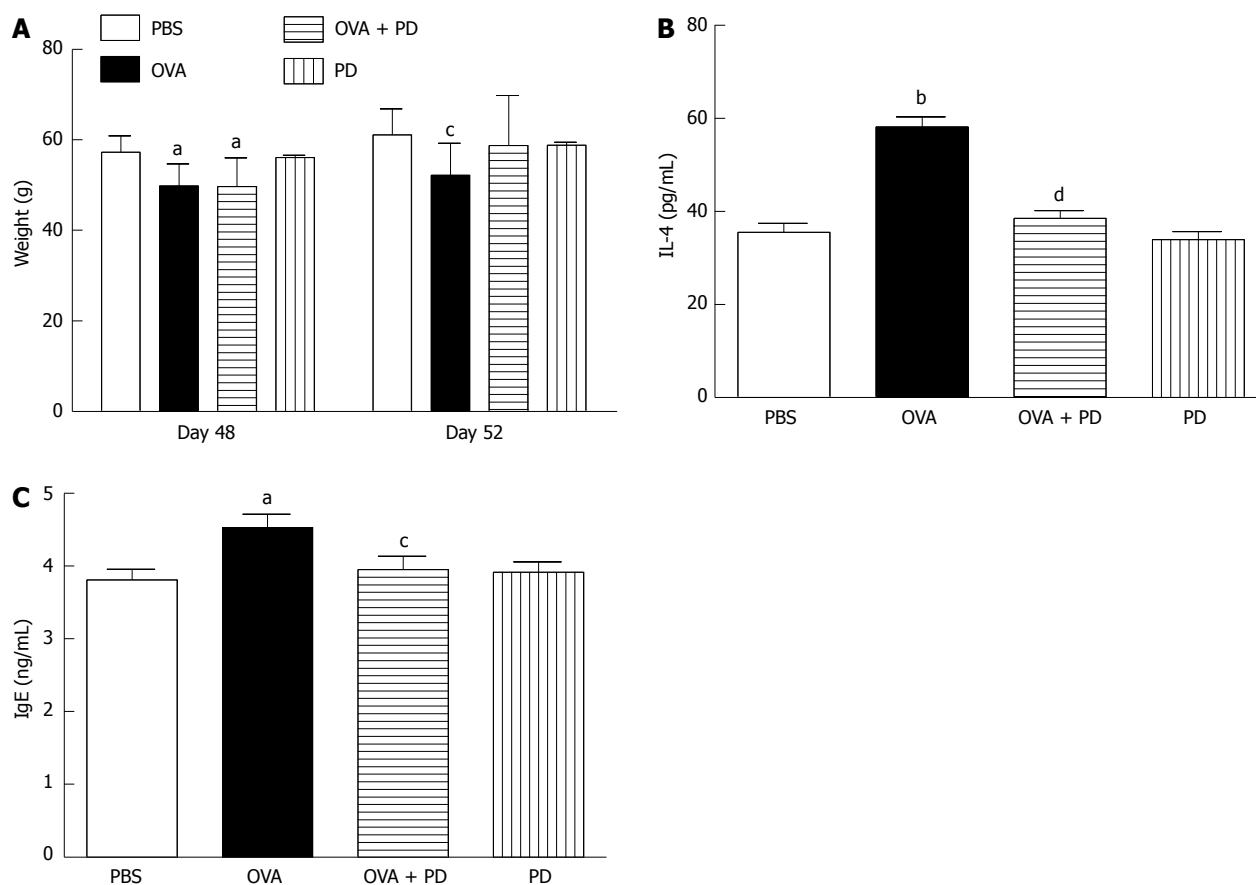


Figure 4 Polydatin suppressed interleukin-4 release and immunoglobulin E production in ovalbumin-allergic rats. A: Body weight of rats on day 48 [left panel, before polydatin (PD) treatment] or day 52 (right panel, after PD treatment) are shown; B: The cytokine levels in rat small intestine tissue or peritoneal lavage solution were analyzed by enzyme-linked immunosorbent assay; C: Statistical analysis of ovalbumin (OVA)-specific immunoglobulin E (IgE) in serum, which were collected from allergic rats administered with or without PD. $n = 8$. ^a $P < 0.05$, ^b $P < 0.01$ vs phosphate-buffered saline (PBS) group; ^c $P < 0.05$, ^d $P < 0.01$ vs OVA group.

analog of PD, on the regulation of intracellular Ca^{2+} signaling has been reported by several groups, although the results vary in different cell types^[29,30]. Furthermore, a previous study in our lab identified PD as a novel mast cell stabilizer in passive cutaneous anaphylaxis mice^[15]. There are two major findings in the present study. Firstly, using an *in vivo* food allergic model, we demonstrated that PD has therapeutic effects against food allergy by decreasing antigen-stimulated mast cell degranulation. Secondly, it was showed that PD suppressed Ca^{2+} mobilization by inhibiting Ca^{2+} entry through SOCs, which were the major contributors to PD-induced mast cell stabilization.

Food allergy is an immunological adverse reaction caused by food, which encompasses a range of disorders including IgE-mediated anaphylaxis, food protein-induced enterocolitis syndrome, and food-induced eosinophilic gastrointestinal disorders. Allergens from eggs seem to be one of the most frequent causes of food allergic reaction reported^[31]. Thus, in this study, we used OVA to sensitize rats and establish a food allergic model. The allergic animal exhibited abnormal intestinal morphology and increased smooth muscle contractility, enhanced Th2 cytokine levels (IL-4), and OVA-specific IgE concentration. Our results are in line with other published data, as IL-4 has been reported to be the hallmark Th2-type

cytokine with multiple immunological functions, including directing Th2 cell differentiation, triggering Ig class switching to IgE in B cells, driving mast cell expansion in intestines^[32], and inducing an exaggerated contractile response in intestinal smooth muscle^[33]. Furthermore, mast cell activation and degranulation, which was due to Ca^{2+} mobilization *via* SOCs, was also demonstrated.

Mast cells are the main effector cells in the pathogenesis of multiple allergic diseases, including asthma, allergic rhinitis, gastrointestinal allergy, and cutaneous anaphylaxis. The majority of mast cell studies have addressed their predominant role in acute allergic reactions (immediate hypersensitivity) and more recently, their roles in late-phase allergic reactions^[34,35]. Therefore, developing new drugs capable of stabilizing mast cells would be valuable for treating diseases attributable to type I hypersensitivity reactions. Using the RBL-2H3 mast cell line, intensive research have been focused on looking for promising drugs to inhibit mast cell activation and degranulation, among which PD showed some latent effects^[15]. In the present study, following administration with PD, the mast cell-dependent food allergic rats did not have apparent anaphylactic symptoms (data not shown) and had a marked decrease in Th2 responsiveness after oral challenge with OVA. We have found that PD significantly reduced IgE

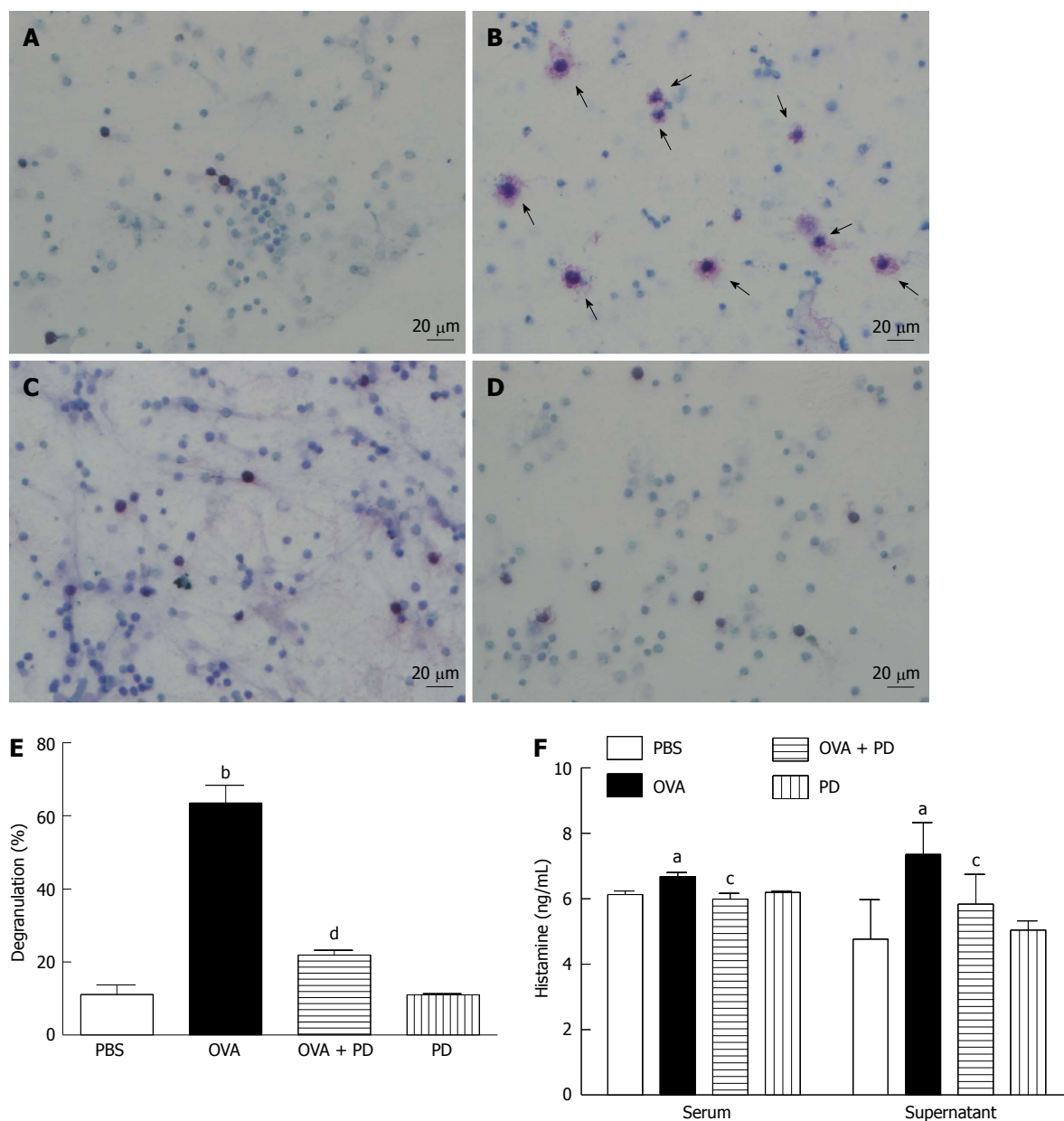


Figure 5 Polydatin therapy reduced mast cell degranulation and activation. A: Phosphate-buffered saline (PBS) group; B: Ovalbumin (OVA) group; C: OVA + polydatin (PD) group; D: PD group; E: Degranulated mast cells were counted from at least 500 total cells and the percentage of degranulation was calculated as degranulated cells against total cells; F: The release of histamine in serum (left panel), rat small intestine tissue, or peritoneal lavage solution (right panel) was measured by enzyme-linked immunosorbent assay. $n = 8$, ^a $P < 0.05$, ^b $P < 0.01$ vs PBS group; ^c $P < 0.05$, ^d $P < 0.01$ vs OVA group. The mast cells in rat small intestine tissue or peritoneal lavage solution were identified by toluidine blue stain. Mast cells were considered degranulated if at least 5 granules appeared outside the cell body. Arrows indicate degranulated mast cells (magnification, $\times 250$).

production by inhibiting Th2 cytokines release. On the other hand, PD showed the potential to block mast cell degranulation by decreasing Ca^{2+} influx *via* SOCs.

The importance of calcium influx in mast cell activation and degranulation has been well recognized^[36]. The degranulation of mast cells is Ca^{2+} dependent, and an increase in intracellular Ca^{2+} characterized by Ca^{2+} entry through SOCs is essential for granules release^[13,27,37]. In this study, we found that PD treatment significantly attenuated FcεRI-elicited intracellular Ca^{2+} increase, indicating that PD stabilized mast cells by suppressing Ca^{2+} mobilization. Furthermore, we found that PD inhibited

Ca^{2+} entry through SOCs. Multiple mechanisms are involved in the regulation of SOC activity. It has recently been discovered that two subunits, STIM1 and Orai1, play a vital role in both the signaling and the permeation mechanisms for Ca^{2+} influx through SOCs. Overexpression of STIM1 together with Orai1 caused a dramatic increase in store-operated Ca^{2+} entry in RBL cells^[38]. The mechanism underlying PD-mediated inhibition of SOCs activity remains unclear.

In summary, the present study established PD as a novel mast cell stabilizer, with the capacity for preventing pathogenesis of food allergy and perhaps other mast cell-

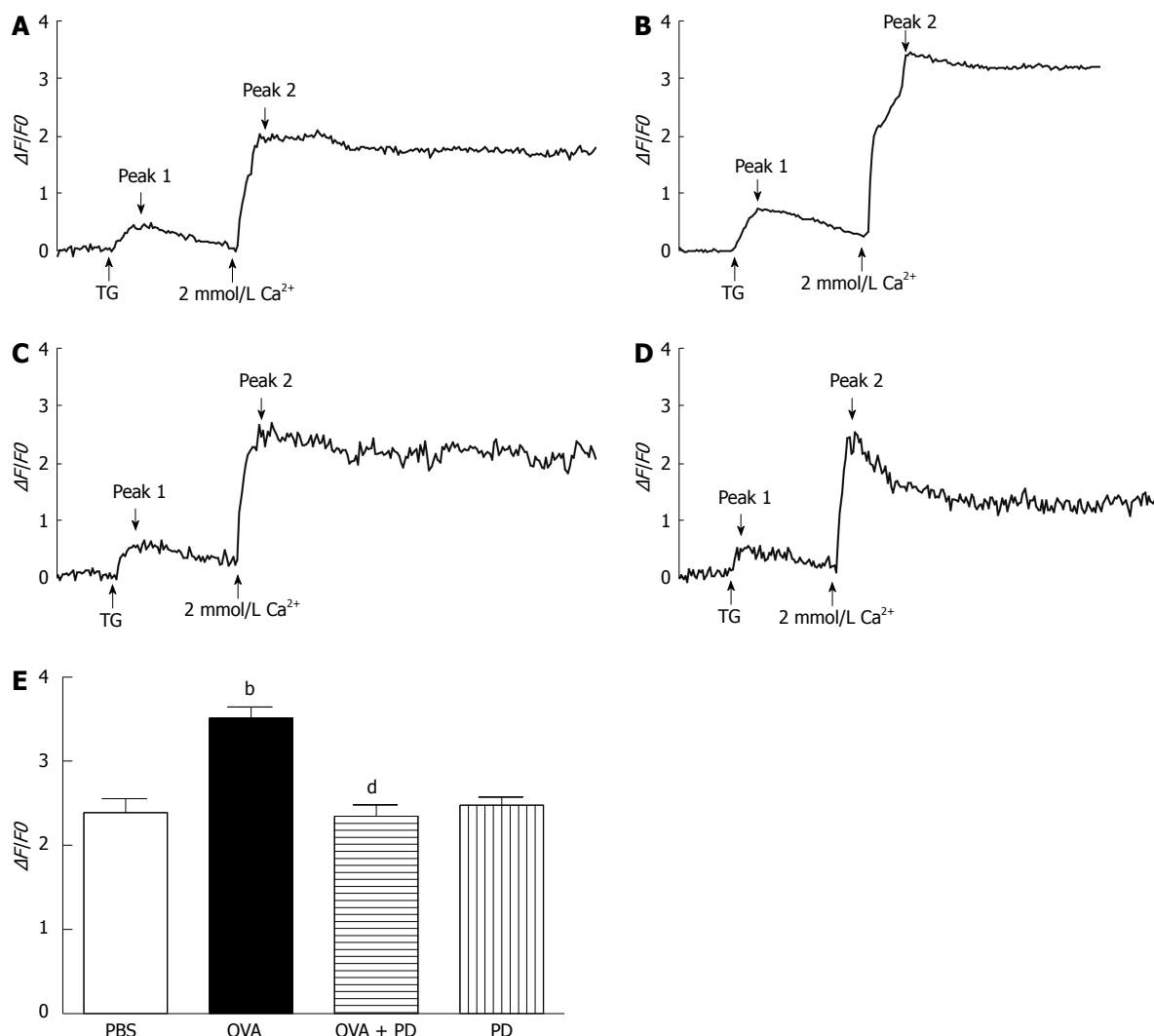


Figure 6 Polydatin reduced Ca²⁺ entry through store-operated calcium channels in food allergic mast cells. Typical responses of thapsigargin (TG)-evoked Ca²⁺ entry through store-operated calcium channels in rat peritoneal mast cells (RPMCs). A: Phosphate-buffered saline (PBS) group; B: Ovalbumin (OVA) group; C: OVA + polydatin (PD) group; D: PD group; E: Averaged peak amplitude of Ca²⁺ entry (second peak) as recorded in each group. ^b*P* < 0.01 vs PBS group; ^d*P* < 0.01 vs OVA group. RPMCs were isolated from Brown-Norway rat treated with or without PD. Intracellular Ca²⁺ was indicated by a fluo-4 fluorescent probe. Total cell numbers are 40-50 for each group, and the cells were from four independent experiments.

dependent allergic diseases through stabilizing mast cells. The underlying mechanism for PD-induced stabilization of mast cells is related to the inhibition of Ca²⁺ mobilization upon FcεRI activation.

COMMENTS

Background

The prevalence of food allergy has increased dramatically during the last three decades, but currently there is no satisfactory therapy except avoidance of the allergen in the diet. However, the offending foods causing an allergic effect are usually essential nutrients to human health. Therefore, to develop a new drug for food allergies is extremely important. Polydatin is a natural component isolated from *Polygonum cuspidatum*, and has been demonstrated to be effective in the treatment of allergic diseases.

Research frontiers

Polydatin is a sort of natural biological material and has been used as a medicine for many diseases. In the area of treatment of allergic diseases with polydatin, the research hotspot is how this product could stabilize mast cells and reduce allergic reactions in passive cutaneous anaphylaxis animals. The

therapeutic effect of polydatin on food allergy has not yet been determined.

Innovations and breakthroughs

In previous studies in other laboratories and author's group, the extract of polydatin has been identified as a novel mast cell stabilizer in passive cutaneous anaphylaxis mice, in addition to its other new therapeutic effects, such as anticancer activity. Using an ovalbumin-sensitization mouse model, the authors are the first group to demonstrate that polydatin could attenuate food allergy by reducing mast cell degranulation. Furthermore, it was shown that polydatin suppressed Ca²⁺ mobilization by inhibiting Ca²⁺ entry through store-operated calcium channels, which were the major contributors to polydatin-induced mast cell stabilization.

Applications

The current results suggest that polydatin is a potential therapeutic drug that could be used in food allergy therapy.

Terminology

Food allergy mediated by immunoglobulin E (IgE) or non-IgE reaction, is an adverse health effect arising from a specific immune response that occurs reproducibly on exposure to a given food. It encompasses a range of disorders including IgE-mediated anaphylaxis, food protein-induced enterocolitis syndrome, and food-induced eosinophilic gastrointestinal disorders; Polydatin, also known as *polygoni cuspidate radix*, is a natural component isolated from *Po-*

lygonum cuspidatum. It has been determined as a resveratrol glucoside with a 3,4,5-trihydroxystilben-3-D-mono-D-glucoside molecular structure. It is traditionally used in South Korea, China, and Japan as a folk remedy for menoxenia, skin burns, gallstones, hepatitis, inflammation, and osteomyelitis.

Peer review

The authors investigated the effect of polydatin, a resveratrol glucoside, on mast cell degranulation and anti-allergic activity. This is a paper with some interesting value.

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